

Supplementary Materials for

HIF-independent synthetic lethality between CDK4/6 inhibition and VHL loss across species

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The PDF file includes:

- Fig. S1. Control experiments for competition experiments done with isogenic ccRCC cell lines treated with CDK4/6 inhibitors.
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- Fig. S3. Changes in proliferation of ccRCC cells after pVHL reconstitution does not account for differential sensitivity to CDK4/6 inhibition.
- Fig. S4. Individual knockdown of CDK4 or CDK6 does not differentially affect the viability of ccRCC cells based on *VHL* status.
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- Fig. S11. Schematic of analogous signaling mechanisms in breast cancer and ccRCC.
- Table S1. sgRNA oligonucleotides.
- Table S2. PCR primers.
- Legends for data files S1 to S3

Other Supplementary Material for this manuscript includes the following:

(available at stke.sciencemag.org/cgi/content/full/12/601/eaay0482/DC1)

Data file S1 (Microsoft Excel format). Results of screen for synthetic lethality with *vhl* inactivation using dsRNA library in *Drosophila* cells.

Data file S2 (Microsoft Excel format). Results of screen for synthetic lethality with *VHL* inactivation using chemical library in human 786-O and UMRC-2 ccRCC cells.

Data file S3 (Microsoft Excel format). Overlap between genes encoding targets of chemicals that scored in chemical screen and human orthologs of *Drosophila* genes that scored in dsRNA screen.

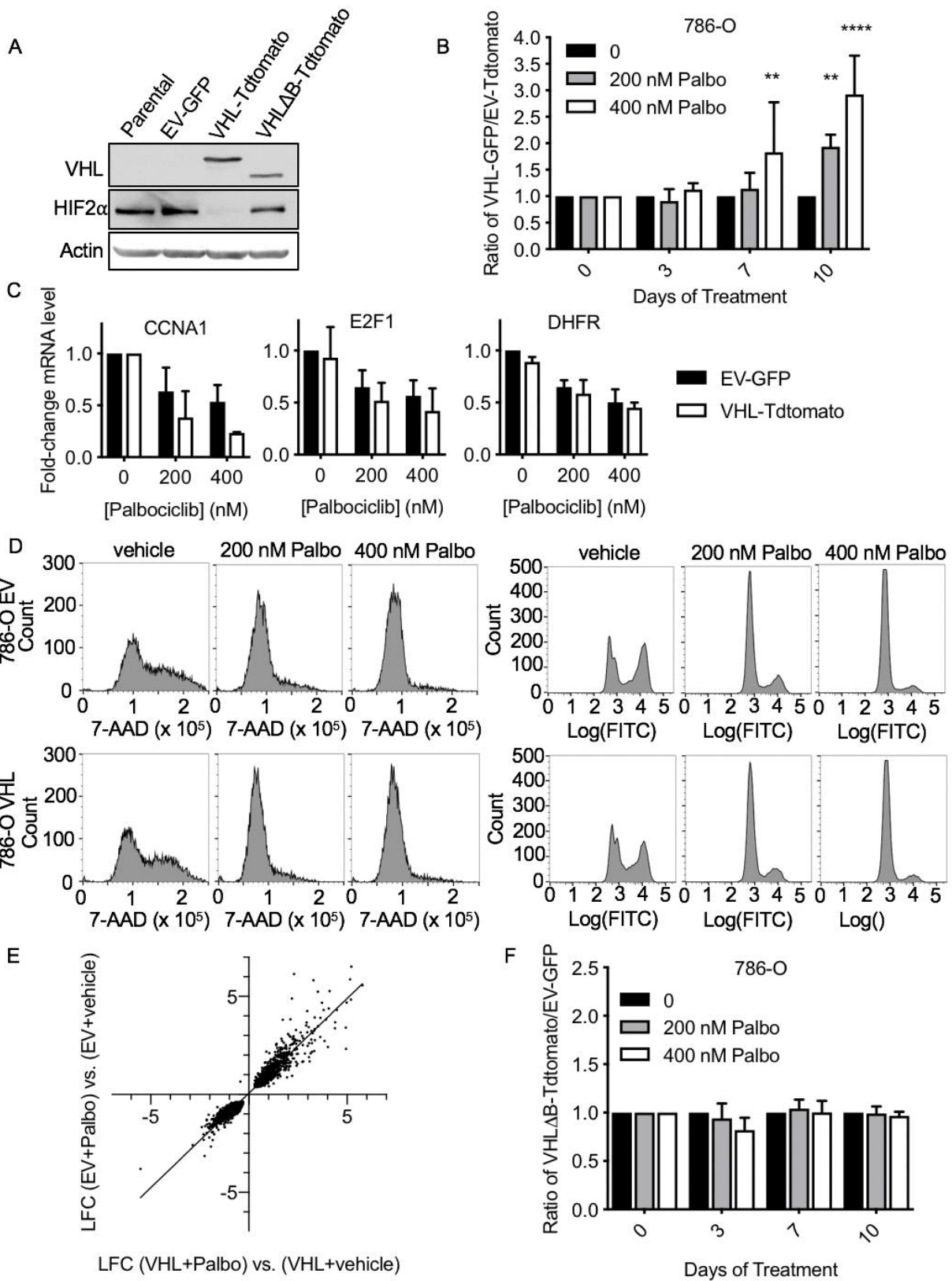


Fig. S1. Control experiments for competition experiments done with isogenic ccRCC cell lines treated with CDK4/6 inhibitors. (A) Immunoblot of *VHL*^{-/-} 786-O cells stably infected, where indicated, with a bicistronic lentivirus expressing *VHL* and *Tdtomato* (VHL-Tdtomato), *GFP* alone (EV-GFP), or a *VHL* variant that lacks the β -domain and *Tdtomato* (VHL Δ B-Tdtomato). The cells expressing the VHL Δ B variant were included for comparative purposes. Blots are representative of 3 experiments. (B) Ratio of VHL-GFP-expressing to EV-Tdtomato-expressing 786-O cells that were mixed 1:1 and then treated with 0, 200, or 400 nM palbociclib for the indicated durations. Data are mean \pm SD from n=3 independent experiments. (C) Relative mRNA expression for *CCNA1*, *E2F1*, and *DHFR* in 786-O cells stably expressing *VHL* or with the empty vector and treated with 0, 200, or 400 nM palbociclib for 48 hours. Data are mean \pm SD from n=2 independent experiments. (D) Flow cytometry analysis after BrdU incorporation and 7-AAD and BrdU-FITC staining of 786-O cells stably infected with lentivirus expressing VHL (786-O VHL) or the empty vector (786-O EV) and treated 0, 200, or 400 nM palbociclib for 24 hours. Data are representative of 3 experiments. (E) Log fold change (LFC) of RNA expression in VHL-Tdtomato and 786-O EV-GFP cells upon treatment with 400 nM Palbociclib or vehicle for 72 hours. (F) Ratio of VHL Δ B-Tdtomato-expressing to EV-GFP-expressing 786-O cells that were mixed 1:1 and then treated with 0, 200, or 400 nM palbociclib for the indicated durations. Data are mean \pm SD from n=3 independent experiments.

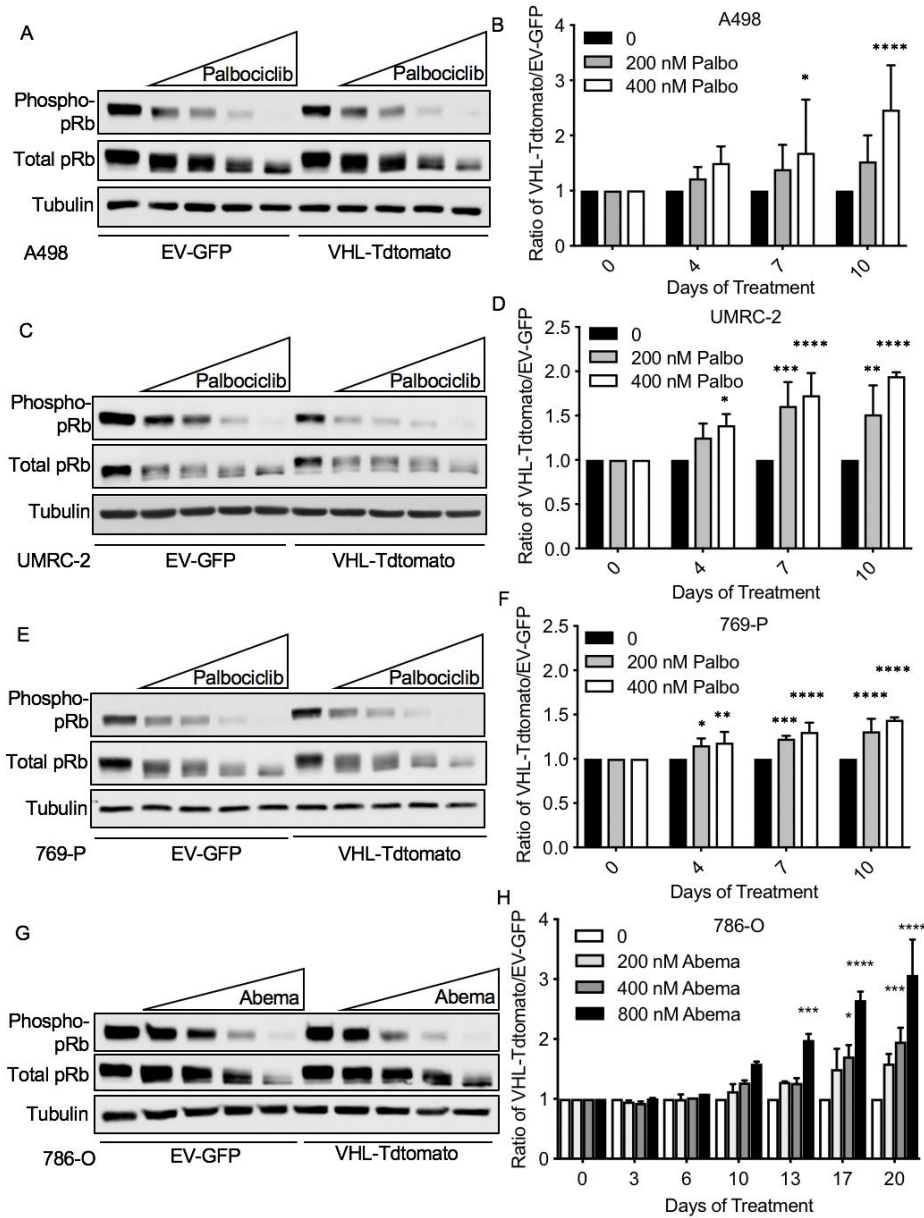


Fig. S2. The CDK4/6 inhibitor palbociclib preferentially inhibits pVHL-deficient cells in various ccRCC cell lines. (A) Immunoblots and densitometry analysis of A498 cells stably expressing *VHL* and *Tdtomato* (VHL-Tdtomato) or *GFP* alone (EV-GFP) and treated with 100, 200, 400, or 800 nM palbociclib (as indicated by the triangle) for 24 hours. Blots are representative of 3 experiments. (B) Ratio of A498 VHL-Tdtomato cells to EV-GFP cells that were mixed 1:1 and then treated with 0, 200, or 400 nM palbociclib for the indicated durations. Data are mean \pm SD of $n=5$ independent experiments. (C and D) As described for (A and B) in UMRC-2 cells. Blots are representative of 3 experiments, and data are mean \pm SD of $n=3$ independent experiments. (E and F) As described for (A and B) in 769-P cells. Blots are representative of 3 experiments, and data are mean \pm SD of $n=3$ independent experiments. (G and H) As described for (A and B) in 786-O cells. Blots are representative of 3 experiments, and data are mean \pm SD of $n=2$ independent experiments. * $P<0.05$, ** $P<0.01$, *** $P<0.001$, and **** $P<0.0001$ by two-way ANOVA tests.

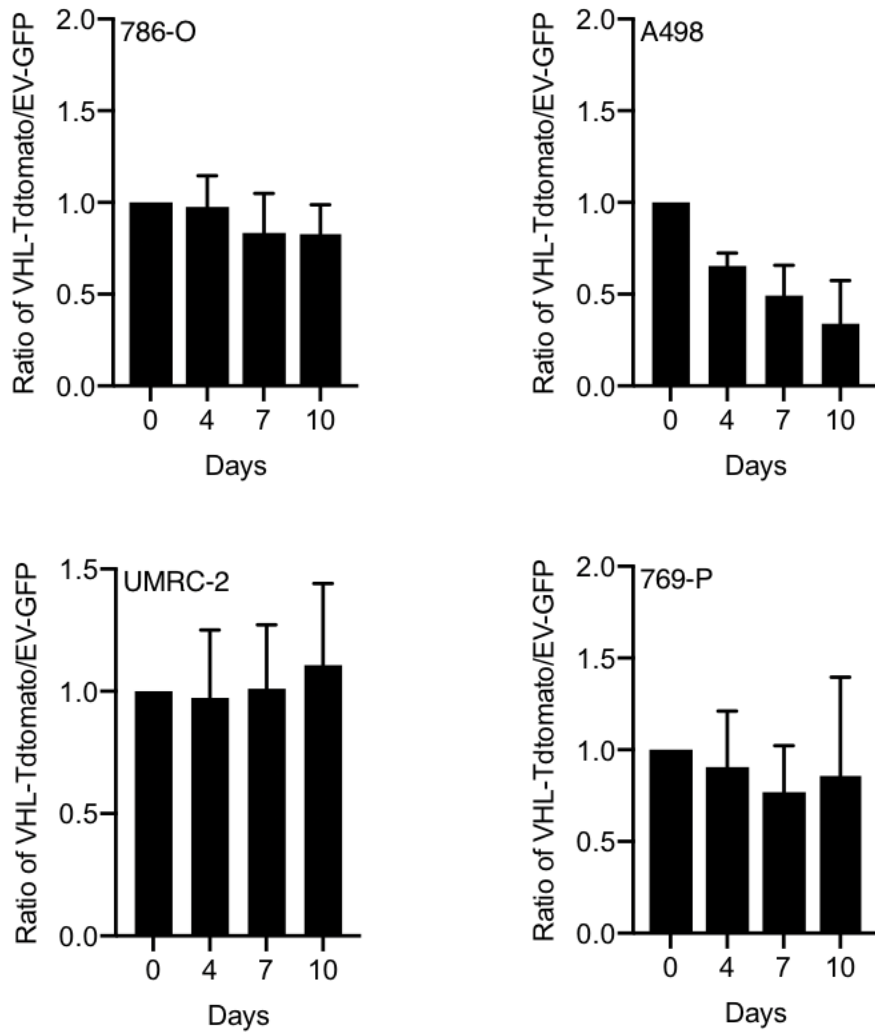


Fig. S3. Changes in proliferation of ccRCC cells after pVHL reconstitution does not account for differential sensitivity to CDK4/6 inhibition. Ratio of 786-O, A498, UMRC-2, and 769-P VHL-Tdtomato cells/ EV-GFP cells over time that were mixed 1:1 on Day 0. Data are mean + SD of n=4 independent experiments.

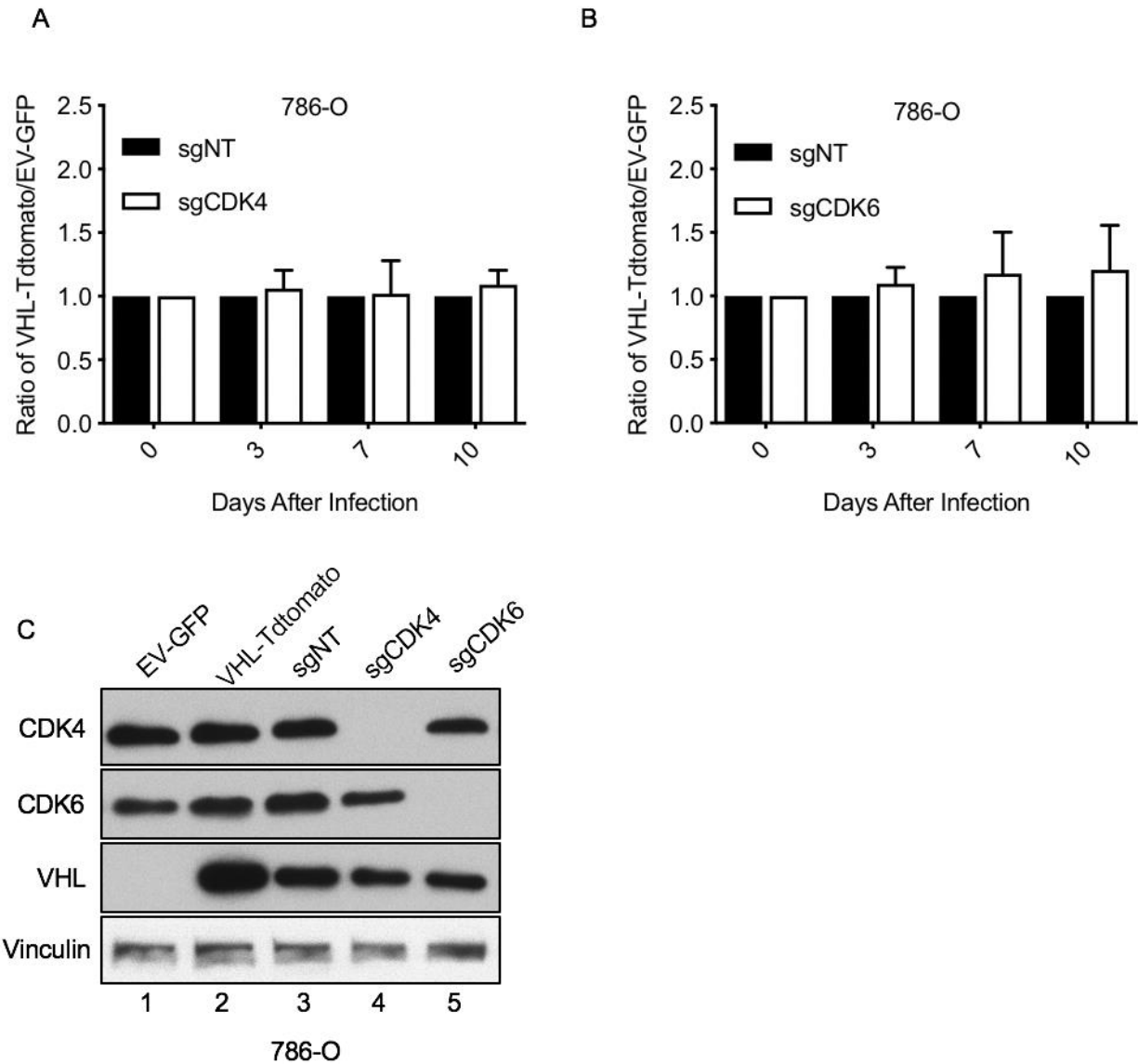


Fig. S4. Individual knockdown of CDK4 or CDK6 does not differentially affect the viability of ccRCC cells based on *VHL* status. (A and B) Ratio of VHL-Tdtomato- or EV-GFP-expressing 786-O cells that were mixed 1:1 and then superinfected with a lentivirus expressing the indicated sgRNAs (sgNT = non-targeting control sgRNA). Data are mean \pm SD of n=6 (A) or 8 (B) independent experiments. (C) Immunoblot of 786-O cells described in (A and B) either prior to (lanes 1 and 2) or 10 days after (lanes 3 to 5) lentiviral infection and mixing. Blot is representative of 3 independent experiments.

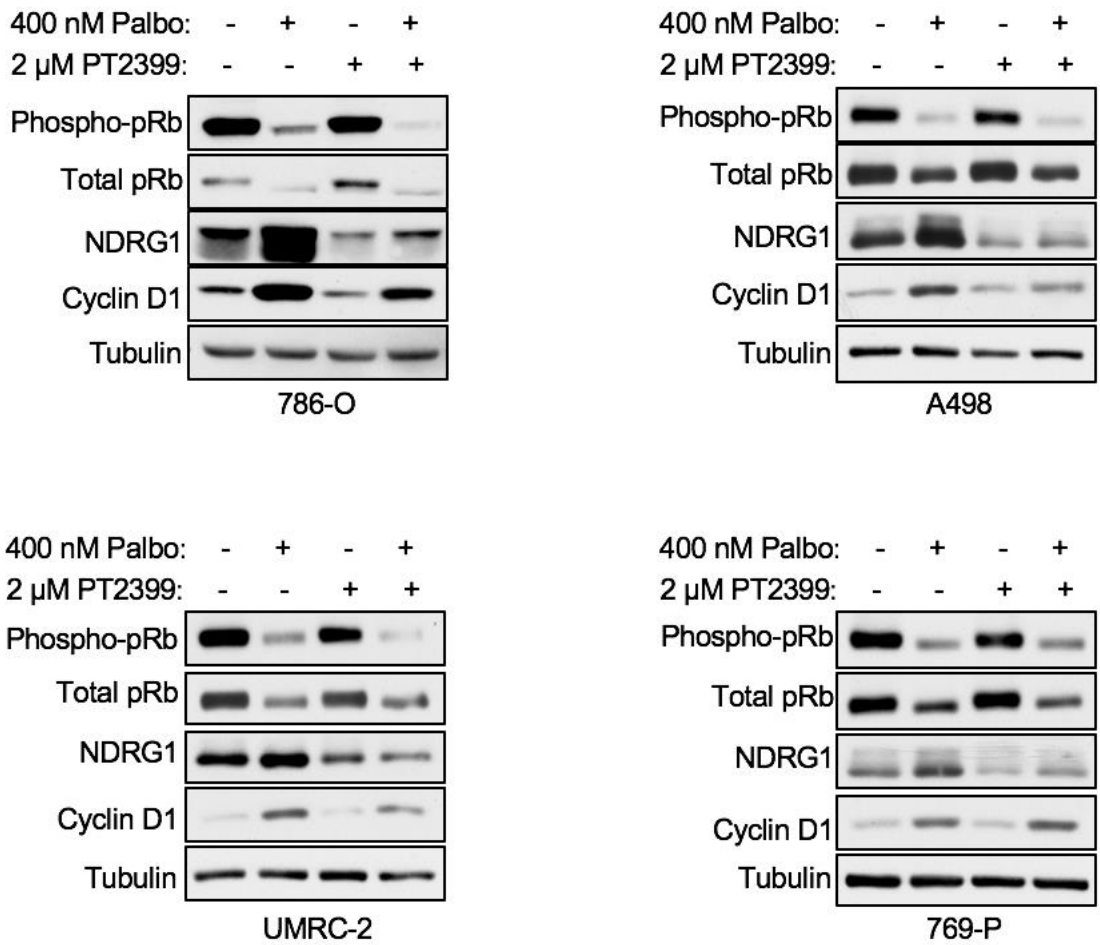
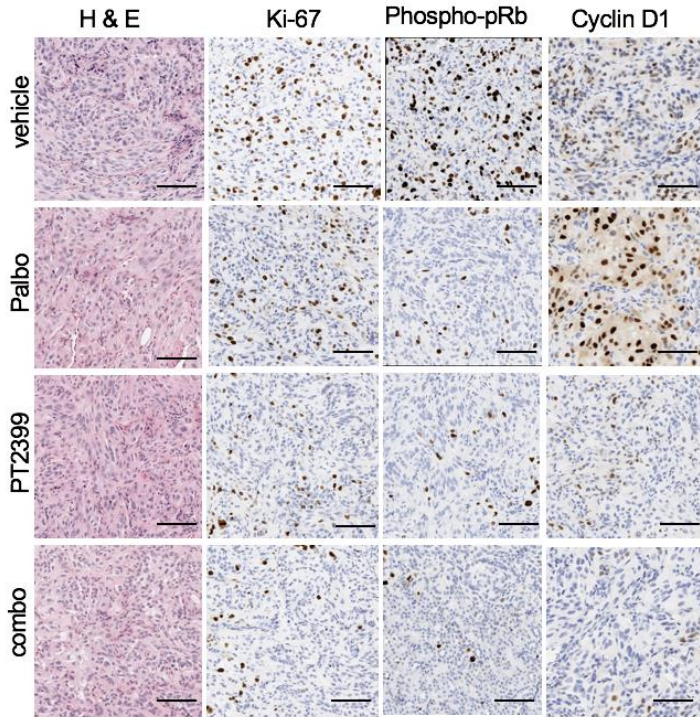
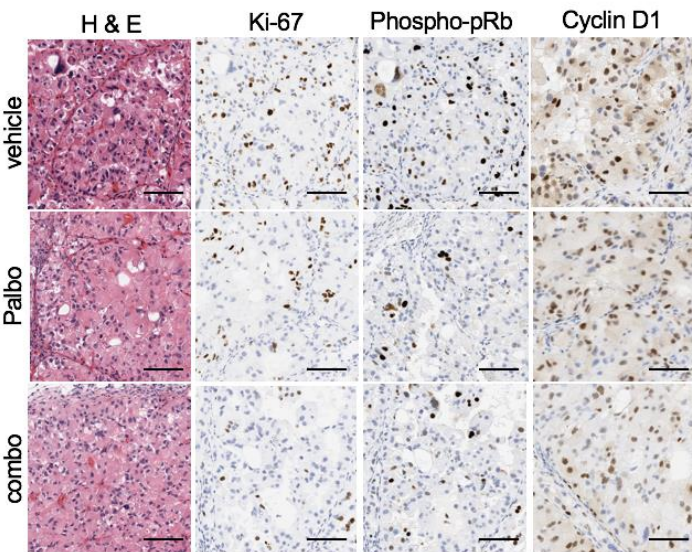


Fig. S5. PT2399 attenuates palbociclib-induced up-regulation of cyclin D1 abundance in HIF-2 α -dependent, but not HIF-2 α -independent, cell lines. Immunoblot of 786-O, A498, UMRC-2, or 769-P EV-GFP cells treated with 2 μ M PT2399, 400 nM Palbociclib, or the combination of both drugs, where indicated for 48 hours. Blots are representative of 3 independent experiments.



786-O



UMRC-2

Fig. S6. Effect of palbociclib, PT2399, and their combination on cyclin D1 and phospho-pRb abundance in vivo. Immunohistochemistry of representative 786-O and UMRC-2 orthotopic tumors isolated from mice treated daily for two days by oral gavage with either vehicle, 65 mg/kg palbociclib (Palbo), 20 mg/kg PT2399 (A only), or both (20 mg/kg PT2399 and 65 mg/kg palbociclib; “combo”). Histology images are representative of n=2 mice for each condition. Scale bars, 100 μ m.

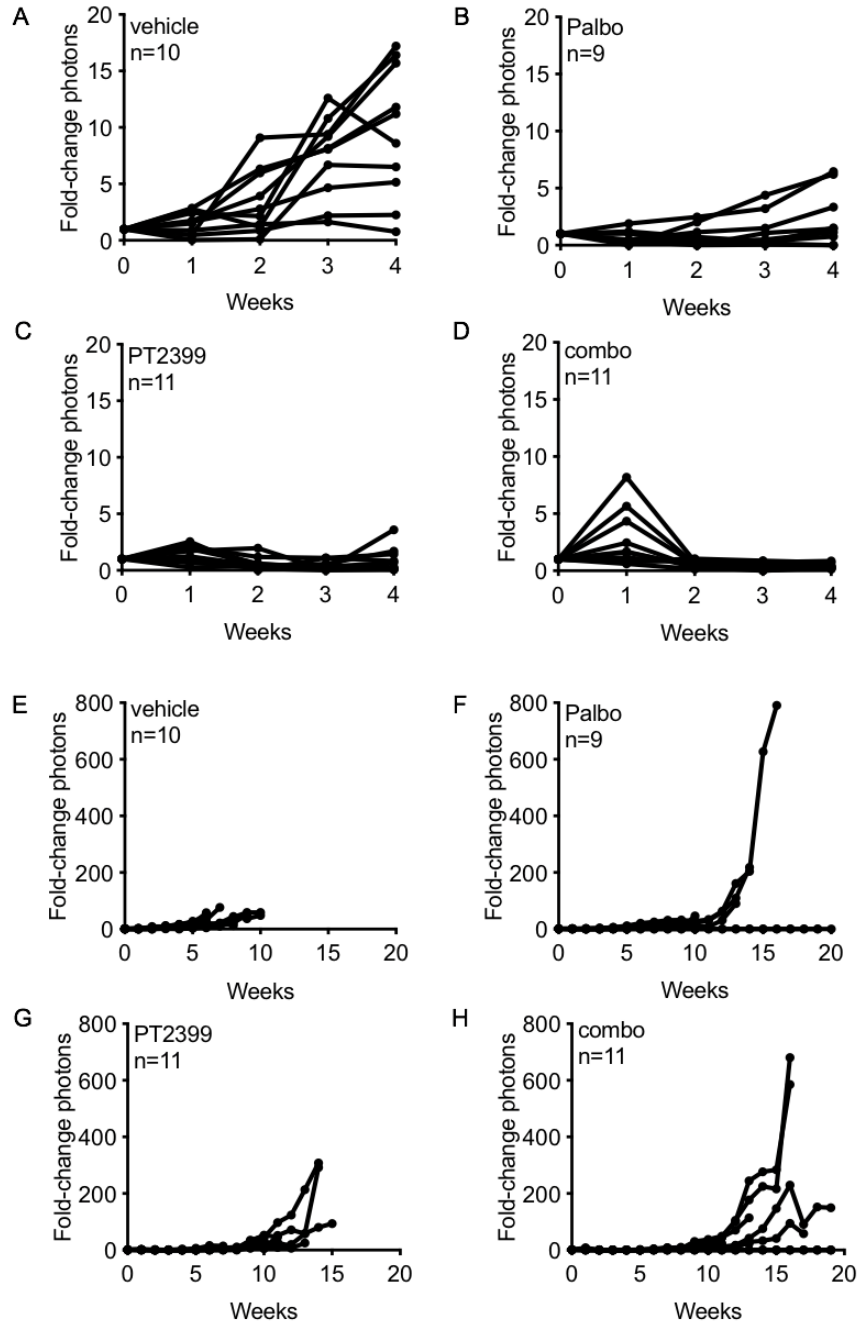


Fig. S7. Growth of ccRCC orthotopic xenografts during treatment with vehicle, palbociclib, PT2399, or their combination. (A to H) Spider plots showing fold-change photons emitted from orthotopic tumors formed by firefly luciferase-expressing 786-O cells as determined by weekly BLI beginning on Day 0 and continuing for 4 weeks (A-D) or up to 20 weeks (E-H) in tumor-bearing mice treated daily for 28 days with vehicle (A and E), 65 mg/kg palbociclib alone (B and F), 20 mg/kg PT2399 alone (C and G), or both (combo; D and H). Each curve represents a different mouse and terminates with the death of that mouse or at 4 and 20 weeks, respectively. At 20 weeks there were 2 mice alive in (F) and 4 mice in (H). The curves for these mice superimpose with the x- axes in these figures.

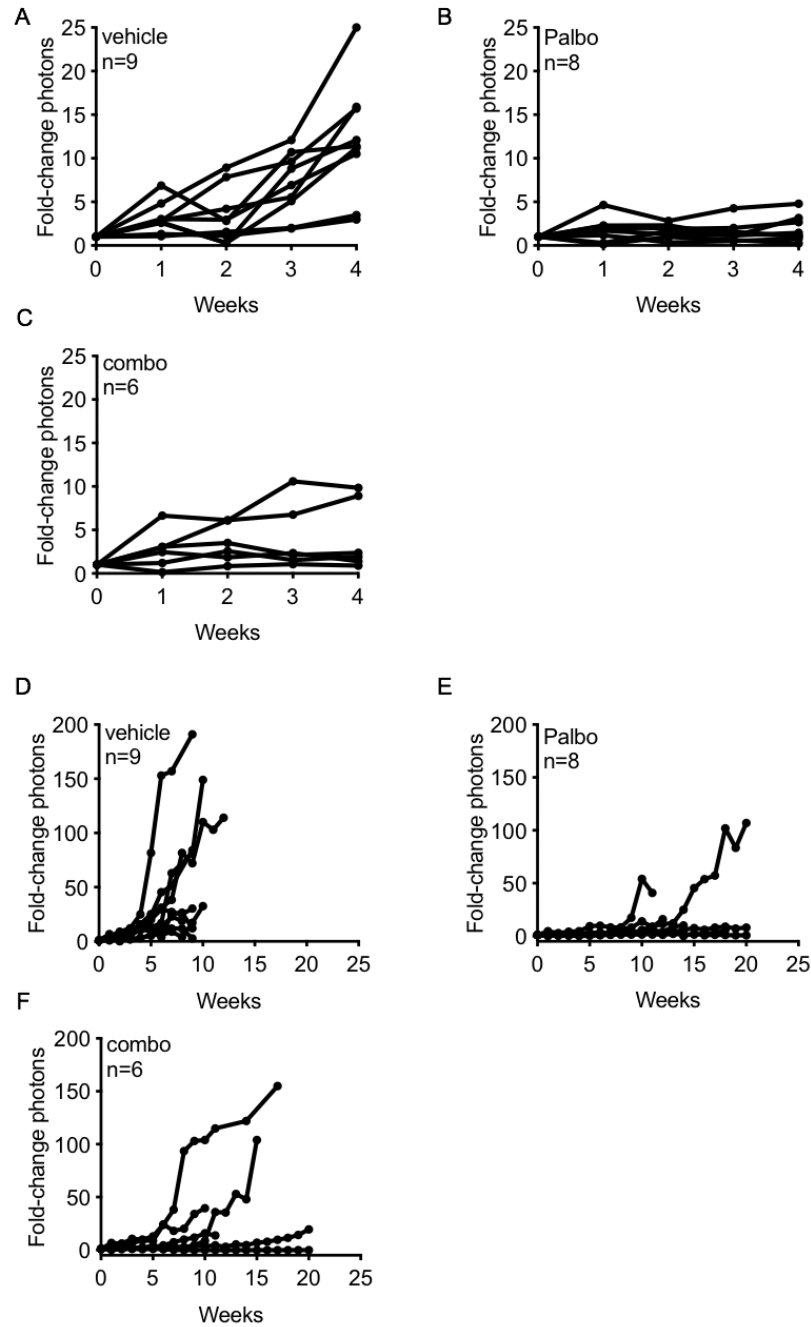


Fig. S8. Growth of HIF-2 α inhibition-resistant *VHL*-null ccRCC orthotopic xenografts during treatment with vehicle, palbociclib, or the combination of palbociclib with PT2399. (A to F) Spider plots showing fold-change in photon emission from orthotopic tumors formed by firefly luciferase-expressing UMRC-2 cells as determined by weekly BLI beginning on day 0 and continuing for 4 weeks (A to C) or 20 weeks (D to F) in tumor-bearing mice treated daily for 4 weeks with vehicle (A and D), 65 mg/kg palbociclib alone (B and E), or both 65 mg/kg palbociclib and 20 mg/kg PT2399 (combo; C and F). Each curve represents a different mouse and terminates with the death of that mouse or at 4 and 20 weeks, respectively. At 20 weeks there were 4 mice alive in (D) and 2 mice in (F). The curves for these mice superimpose with the x- axes in these figures.

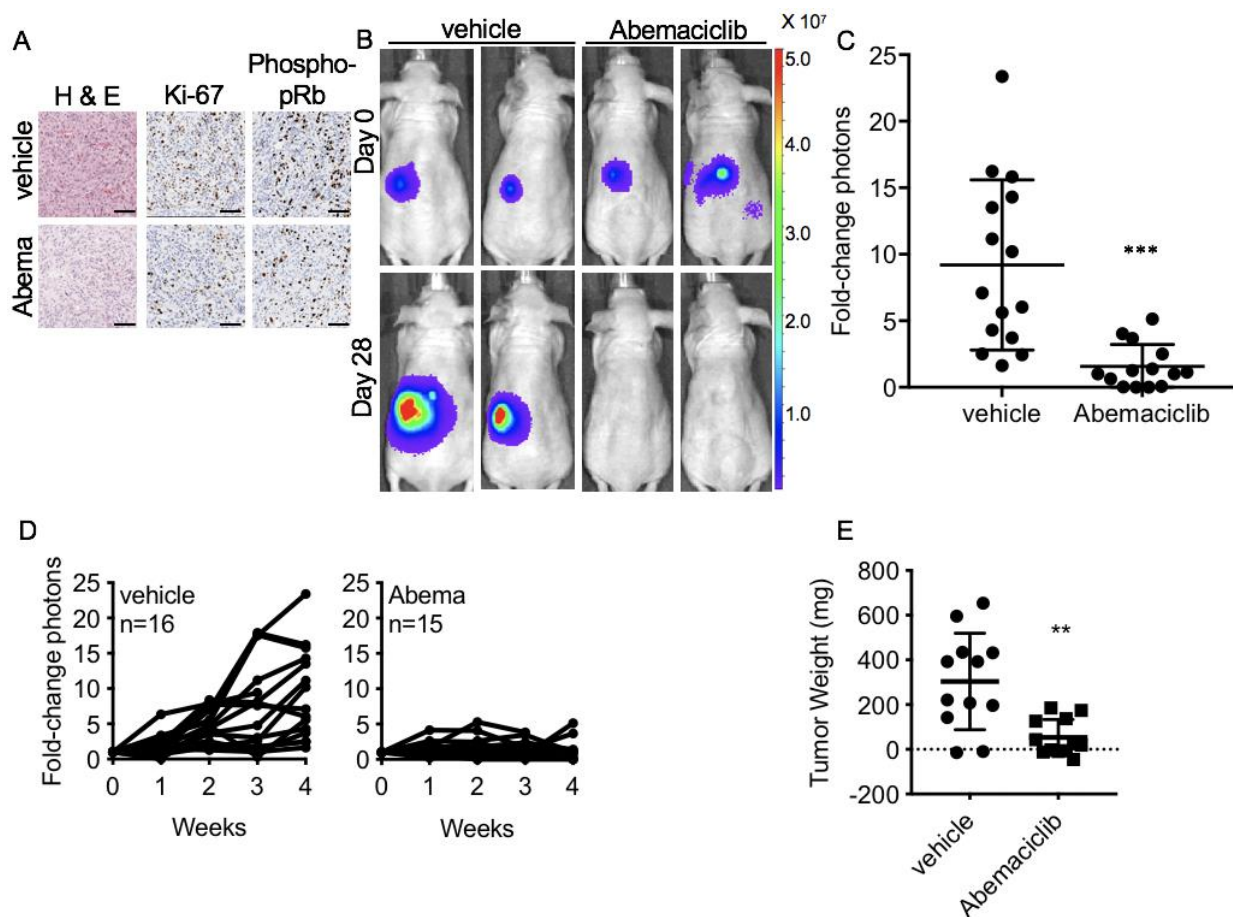


Fig. S9. Antitumor activity of abemaciclib in *VHL*-null ccRCC orthotopic xenografts. (A) Immunohistochemistry of 786-O orthotopic tumors treated with vehicle or 60 mg/kg abemaciclib once daily for two days. Histology images are representative of $n=2$ mice for each condition. Scale bars, 100 μm . (B) Representative BLI of orthotopic tumors formed by firefly luciferase-expressing 786-O cells before and after vehicle or 60 mg/kg abemaciclib dosed daily by oral gavage for 28 days ($n=15$ or 14 mice from two independent experiments, respectively). (C) Quantification of BLI for mice described in (B). Photon measures on day 28 were normalized to those on day 0 for each mouse individually ($p=0.0004$ by Welch's t-test). (D) Spider plots showing fold-change in photon emission from tumor-bearing mice described in (B) as determined by weekly BLI over the course of 28 days of treatment with vehicle (left) or abemaciclib (Abema; right). (E) Tumor weights from mice described in (B). $n=12$ mice for vehicle, 11 for abemaciclib; $**P<0.001$ ($P=0.0020$) by Welch's t-test.

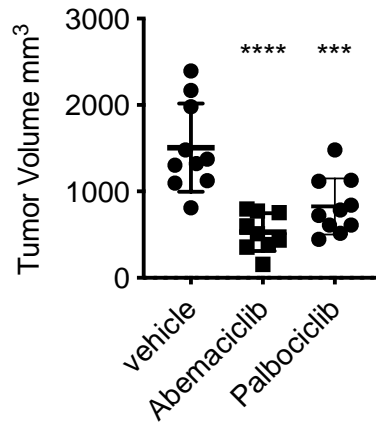


Fig. S10. Antitumor activity of palbociclib and abemaciclib in a ccRCC PDX model.

Volume of patient-derived xenograft tumors after 25 days of treatment with vehicle, 75 mg/kg palbociclib daily, or 60 mg/kg abemaciclib twice daily. n=10 mice for vehicle, 9 for abemaciclib, and 10 for palbociclib; *** $P < 0.001$, **** $P < 0.0001$ by one-way ANOVA.

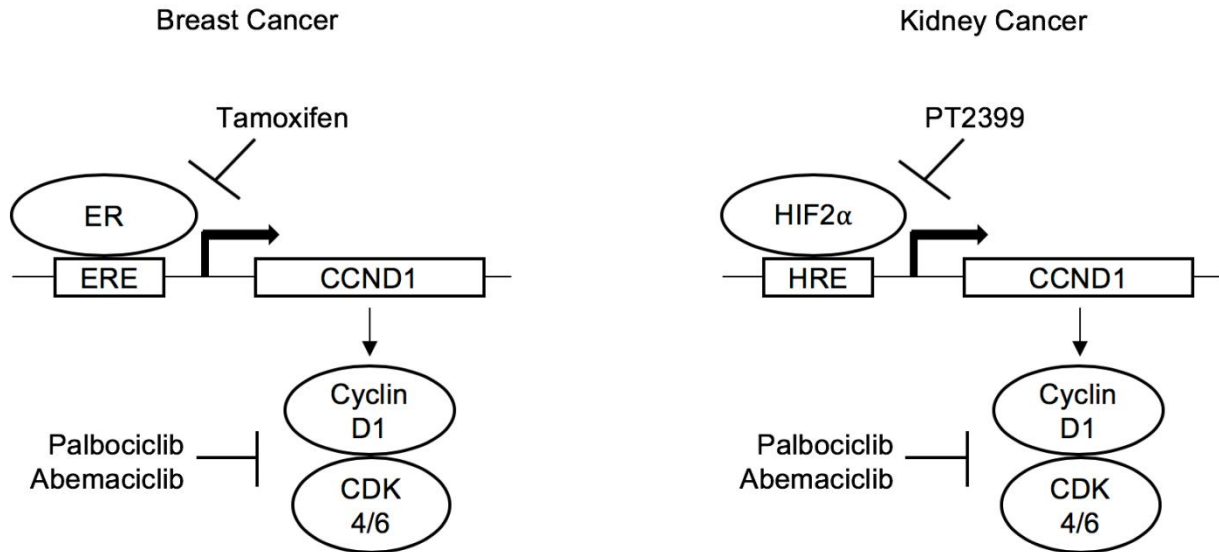


Fig. S11. Schematic of analogous signaling mechanisms in breast cancer and ccRCC.

The estrogen receptor (ER) and HIF2 α bind to an estrogen response element (ERE) and hypoxia-response element (HRE), respectively, that control the transcription of *CCND1*, which encodes Cyclin D1. In breast cancer ER is a major regulator of *CCND1* and ER antagonists (e.g. tamoxifen) therefore downregulate Cyclin D1. Similarly, HIF2 α regulates *CCND1* in ccRCC and HIF2 α antagonists (e.g. PT2399) can downregulate Cyclin D1 in HIF2 α -dependent ccRCC lines. Adding a CDK4/6 inhibitor enhances the ability of ER antagonists to control breast cancer proliferation, presumably because they both directly or indirectly converge on the activity of CDK4/6. By analogy, adding a CDK4/6 inhibitor might enhance the activity of a HIF2 α inhibitor in ccRCC.

Table S1. sgRNA oligonucleotides. The sequences of the sgRNA oligonucleotides used for editing (including BsmBI/Esp3I overhangs) are listed in the table.

<i>sgRNA oligonucleotide target/name</i>	<i>Sequence</i>
<i>CDK4</i> sg1 sense	5'-CACCGGTCCACATATGCAACACCTG-3'
<i>CDK4</i> sg1 antisense	5'-AAACCAGGTGTTGCATATGTGGACC-3'
<i>CDK4</i> sg2 sense	5'-CACCGGTCTACATGCTCAAACACCA-3'
<i>CDK4</i> sg2 antisense	5'-AAACTGGTGTGTTGAGCATGTAGACC-3'
<i>CDK6</i> sg1 sense	5'-CACCGCCAGCAGTACGAATGCGTGG-3'
<i>CDK6</i> sg1 antisense	5'-AAACCCACGCATTCGTAAGTACTGCTGGC-3'
<i>CDK6</i> sg2 sense	5'-CACCGTGACCAGCAGTACGAATGCG-3'
<i>CDK6</i> sg2 antisense	5'-AAACCGCATTCGTAAGTACTGCTGGTCAC-3'
Non-targeting sgRNA sense	5'-CACCGGGAGGCTAAGCGTCGCAA-3'
Non-targeting sgRNA antisense	5'-AAACTTGCGACGCTTAGCCTCCC-3'
<i>RB1</i> sense	5'-CACCGCGGTGGCGGCCGTTTTTCGG-3'
<i>RB1</i> antisense	5'-CCGAAAAACGGCCGCCACCGCGGTG-3'
<i>EPAS1</i> sense	5'-CACCGTCATGAGGATGAAGTGCA-3'
<i>EPAS1</i> antisense	5'-AAACTGCACTTCATCCTCATGAC-3'

Table S2. PCR primers. The sequences of the PCR primers used in this study are listed in the table.

Primer target/name	Sequence
<i>Drosophila sima</i> forward	5'-TTTGCCATTGAAAACCGACGA-3'
<i>Drosophila sima</i> reverse	5'-CTTGAGGAAAGCGATGGTGAT-3'
<i>Drosophila globin</i> forward	5'-GAAGGTACCGCATAAACATGAACAGCGATGAGG-3'
<i>Drosophila globin</i> reverse	5'-GAAGGTACCGCTGCCTCATCTACTTGGCGTTG-3'
<i>Drosophila LDH</i> forward	5'-GTCTGTTGGCCAGTTGCTGAGG-3'
<i>Drosophila LDH</i> reverse	5'-CTGGACATCGGACATGATGTTGGCGGAC-3'
<i>Drosophila CG11652</i> forward	5'-GTTGGACGTGATCCAACCCAGCAG-3'
<i>Drosophila CG11652</i> reverse	5'-GATCGTCCAGAGCCGCTGCCTTG-3'
<i>Drosophila act5c</i> forward	5'-GAGCGCAAGTACTGTGTCTGG-3'
<i>Drosophila act5c</i> reverse	5'-GACTCGTCGTA CTCTGCTGG-3'
Human <i>CCNA1</i> forward	5'-GCACCCTGCTCGTCACTTG-3'
Human <i>CCNA1</i> reverse	5'-CAGCCCCAATAAAAGATCCA-3'
Human <i>E2F1</i> forward	5'-CTTCGTAGCATTGCAGACCC-3'
Human <i>E2F1</i> reverse	5'-TATGGTGGCAGAGTCAGTGG-3'
Human <i>DHFR</i> forward	5'-GGTCTGGATAGTTGGTGGCA-3'
Human <i>DHFR</i> reverse	5'-AACACCTGGGTATTCTGGCA-3'
Human <i>ACTB</i> forward	5'-ACCAACTGGGACGACATGGAGAAA-3'
Human <i>ACTB</i> reverse	5'-TAGCACAGCCTGGATAGCAACGTA-3'
Human <i>CCND1</i> forward	5'-CCGTCCATGCGGAAGATC-3'
Human <i>CCND1</i> reverse	5'-ATGGCCAGCGGGAAGAC-3'

Data file S1. Results of screen for synthetic lethality with vhl inactivation using dsRNA library in *Drosophila* cells. This data file contains the list of dsRNAs against *Drosophila* genes, the ID of the specific dsRNA amplicons tested, and the Z-scores for the effects of each dsRNA on S2R+ cells that have (WT) or lack (vhl-null) vhl, the *Drosophila* ortholog of the human *VHL* tumor suppressor gene.

Data file S2. Results of screen for synthetic lethality with *VHL* inactivation using chemical library in human 786-O and UMRC-2 ccRCC cells. This data file contains the map of each small molecule library plate, as well as the Z-scores for the effect of each small molecule at each concentration on cellular fitness (as measured by fluorescence of GFP-expressing cells) of 786-O and UMRC-2 cells that lack functional *VHL* (EV) or that have been reconstituted to express functional *VHL* (VHL).

Data file S3. Overlap between genes encoding targets of chemicals that scored in chemical screen and human orthologs of *Drosophila* genes that scored in dsRNA screen. This data file contains the genes in each portion of the venn diagram in Figure 2D.